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ZOO-CHEMICAL PROFILE, ZOO-SYNTHESIS OF METALLIC NANOPARTICLES AND THEIR APPLICATIONS: A SHORT REVIEW

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ABSTRACT

Zoochemicals refer to animal chemical composition, including secondary metabolites, that is similar to plant and microbial secondary metabolites. Zoochemicals are naturally reducing and capping agents that respond to the zoosynthesis of metallic nanoparticles and promising nano-based drugs. The term "zoosynthesis of metallic nanoparticles" refers to zoo-extracts from animal sources that contain secondary metabolites that are involved in metal ion reducing and capping agents that form metal atoms. These are called zoo-synthesized metallic nanoparticles. Animal secondary metabolites like alkaloids, terpenoids, saponins, etc. are produced by marine invertebrates, which are equivalent to plant secondary metabolites and have been used in environmental and biomedical applications. In the present review, we focused on and discussed zoochemicals of invertebrate origin and their applications. Alkaloids, terpenoids, and saponins were major secondary metabolic zoocompounds observed in most of the marine invertebrate phylum, such as Porifera, Mollusca, and Echinodermata, with a suitable solvent extracted, and zoo-extract was used as a potential biological activity, such as antioxidant, anti-inflammatory, cytotoxic, insecticide, and antimicrobial, while also metallic nanoparticle synthesis, such as CuO, ZnO, Ag, and Au nanoparticles. In the present review, a succession of zoochemicals from marine invertebrates are effective, eco-friendly drugs and have biological activity in the search for new drugs, while zoochemicals are evidence of metal-reducing agents that use animal-mediated nanoparticle synthesis, which results in effective, eco-friendly nanodrugs.

Keywords: Zoochemicals, Nanoparticles synthesis, Biological activity, Marine invertebrate.

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INTRODUCTION

Zoochemicals refer to animal chemical composition, including secondary metabolites, that is similar to plant and microbial secondary metabolites [1,2,3]. The present short review focuses on zoochemicals

accruing from animal sources, including marine invertebrates that provide valuable secondary metabolites and metal-reducing agents in response to the zoosynthesis of metallic nanoparticles. We observed zoochemicals from most of the invertebrate

phylum, including some notable phylum such as Porifera, Mollusca, and Echinodermata. Tables 1 and 2 show the biological and nanoparticle synthesis of animal sources.

Phylum Porifera zoochemicals

Hyattella intestinalis marine sponge 70% ethanolic zoo-extract contained 60 zoochemicals were identified using GC-MS techniques [2], and tryptamine alkaloids were confirmed using HPLC techniques [4].

Luffariella herdmani marine sponge with extraction of dichloromethane and methanol with zoochemicals contained alkaloids, terpenoids, sterols, saponins, and anthraquinones, by Kuruppuarachchi and Gunathilake [5].

Hyrtios erectus, Pseudoceratina purpurea, Carteriospongia foliascens, Stylissa carteri, and Axinella acanthelloides marine sponges extracted with ethanol, methanol, acetone, hexane, and chloroform were screened for zoo-chemical profiles, while the presence of alkaloids and terpenoids in most of the extract and sponges was observed by Muthiyan et al. [6].

GC-MS analysis of marine sponges Haliclona (Gellius) sp., Lamellodysidea herbacea, and Spheciospongia inconstans identified mainly sterols and fatty acids. Haliclona (Gellius) sp. showed 23 compounds were identified, Lamellodysidea herbacea had 21 compounds identified, and Spheciospongia inconstans had 19 compounds identified using GC-MS, while the zoochemicals of alkaloids, saponins, and terpenoids were qualified in all three marine sponges [7].

Tetilla dactyloidea marine sponge extract with methanol, and six compounds were identified using GC-MS, including 9-Octadecenoic acid (Z)-, 2-hydroxy-1-(hydroxymethyl) ethyl ester, by Krishnan et al. [8].

Haliclona sp., Dactylospongia elegans, Aaptos suberitoides, and Stylissa massa zoochemicals of alkaloids, saponins, flavonoids, and tannins were present, according to Rivera and Uy, [9].

Phylum Mollusca zoochemicals

Sea hare *Dolabella auricularia* revealed the zoochemicals contained in alkaloids, saponins, steroids, tannins, and terpeneoids, by Tayone *et al.* [10].

Phylum Echinodermata zoochemicals

Sea urchin Salmacis virgulata test zoochemicals contained 59 identified zoochemicals, including hexadecanoic acid and stigmasterol, while one unknown zoochemicals contained 60 zoochemicals, using GC-MS techniques by Karnan et al. [11].

Sea urchin *Diadema setosum* gonad with an extract of methanol, ethyl acetate, and n-hexane fraction was determined to be zoochemically alkaloid, saponin, and phenolic by Rompas *et al.* [12].

Sea cucumber *Bohadschia vitiensis* zoochemicals, which contained saponins, terpenoids, and sterols, by Jayathilake and Gunathilake, [13].

The zoochemical composition of selected sea stars *Linckia laevigata*, *Protoreaster nodosus*, and *Acanthaster planci* showed the presence of alkaloids, flavonoids, saponins, triterpenoids, and cardiac glycosides while being rich in saponins and steroids [14].

The isolated compounds are holothurin A (1) and echinoside A (2) triterpene saponins from sea cucumber *Pearsonothuria graeffei*, which were separated by HPLC [15].

Twenty one lanostane-type nonsulphated triterpene glycosides were isolated from *Bohadschia cousteaui* (sea cucumber) body walls. 10 new saponins, including 2 pentasaccharide, 8 hexasaccharide saponins and 11 known triterpene glycosides, were isolated by HPLC and their structures determined by NMR techniques [16].

Table 1: Invertebrate animal with Biological activity

Invertebrate animal			
Phylum	Study animals	Activity	References
Echinodermata	Sea urchin Salmacis virgulata test	In silico antifungal	Karnan et al. [11]
Porifera	Marine sponge Hyattella intestinalis	Insecticidal	Karnan et al. [17]
Porifera	Marine sponge Luffariella herdmani	In vivo toxicity, in vitro anti-inflammatory	Kuruppuarachchi and Gunathilake, [5].
Cnidaria	Jellyfish Aurelia aurita	Antioxidant	Khalil et al. [18].
Mollusca	Marine snail Hemifusus colosseus	Antimicrobial and antioxidant	Nguyen et al. [19].
Echinodermata	Sea urchin <i>Diadema</i> setosum	Antibacterial	Rompas <i>et al.</i> [12].
Porifera	Marine sponge <i>Hyrtios</i> erectus	Cytotoxic	Muthiyan et al. [6].
Echinodermata	Sea cucumber Bohadschia vitiensis	Anti-inflammatory, Antioxidant and Toxicity	Jayathilake and Gunathilake, [13].
Mollusca	Egg strings from sea hare Dolabella auricularia	Antioxidant	Tayone et al. [10].
Arthropods	Ground beetle	Antibacterial	Yahaya et al. [20].
Porifera	Marine sponges Haliclona sp., Lamellodysidea herbacea, and Spheciospongia inconstans,	Antimicrobial and cytotoxicity	Putra and Hadi, [7].
Porifera	Marine sponge Tetilla dactyloidea	Anticancer	Krishnan et al. [8].
Arthropods	Marine blue swimmer crab (Portunus pelagicus) and mud crab (Scylla tranquebarica)	Antibacterial	Laith et al. [21].
Cnidaria	Jellyfish Porpita porpita	Antimicrobial	Umamageswari <i>et al.</i> [22]
Arthropods	Soft-shelled and hard-shell crab <i>Charybdis lucifera</i>	Antioxidant activity	Soundarapandian <i>et al.</i> [23]
Echinodermata	Sea cucumber Bohadschia cousteaui	Antimicrobial	Elbandy et al. [16]
Porifera	Marine sponges Haliclona sp., Dactylospongia elegans, Aaptos suberitoides and Stylissa massa	Antioxidant and cytotoxic	Rivera and Uy, [9]

Zoo-synthesis of metallic nanoparticles using Invertebrate animals

The term "zoosynthesis of metallic nanoparticles" refers to zoo-extracts from animal sources that are involved in metal ion (M⁺) reducing and capping agents, which form

metal atom (M^0) . These are called zoosynthesized metallic nanoparticles with zooextracts that contain animal secondary metabolites. Table 2 shows the metallic nanoparticle synthesis of animal sources, called zoo-synthesized NPs.

Table 2: Invertebrate animal with metallic nanoparticle synthesis

Invertebrate	Metallic NPs	Application	References
Sea urchin Salmacis virgulata test	CuONPs	-	Karnan et al. [11]
Marine sponge Hyattella intestinalis	ZnONPs	Insecticidal activity	Karnan et al. [1]
Marine sponge Hyattella intestinalis	CuONPs	Insecticidal activity	Karnan <i>et al.</i> [17,4]
Brittle star Ophiocoma scolopendrina	AgNPs	Antibacterial, antioxidant, anti-diabetic and catalytic degradation of organic dyes	George et al. [24]
Marine sponge Spongia officinalis	ZnONPs	Antimicrobial and insecticidal activity	Hasaballah et al. [25]
Marine Bivalvia shell	AgNPs	Larvicidal activity	Sivanandham Velavan et al. [26]
Marine sponge Haliclona	AgNPs	-	Hamed et al. [27]
Marine sponge Acanthella elongata	AgNPs	-	Inbakandan et al. [28]
Marine sponge Acanthella elongata	AuNPs	-	Inbakandan et al. [29]

CONCLUSION

Alkaloids, terpenoids, and saponins are major secondary metabolic zoocompounds observed in most invertebrate animals and used for potential biological activity and zoo-mediated metallic nanoparticle synthesis. In the present review, a succession of zoochemicals are effective, ecofriendly drugs, and there is evidence of metal-reducing agents that use animal-mediated nanoparticle synthesis, which results in effective nanobased, eco-friendly drugs.

REFERENCES

- Karnan R., Velavan S., Mariappan P., & Sukumaran M. (2023c). Evaluation of Insecticidal Activity of Zoochemical-Assisted Zinc Oxide Nanoparticle Using Marine Invertebrate Hyattella intestinalis (Lamarck, 1814). Uttar Pradesh Journal of Zoology, 44(21), 31–39.
- Karnan R, Sukumaran M, Velavan S. (2022). Extraction and identification of zoochemicals in marine sponge *Hyattella intestinalis* (Lamarck, 1814) (Phylum: Porifera) using GC-MS technique. Intern. J. Zool. Invest. 8(Special Issue): 113-118.

- 3. Lindshield B. (2018). Kansas State University Human Nutrition (FNDH 400) Flexbook. New Prairie Press, Kansas State University Libraries.
- Karnan, R., Sukumaran, M., & Velavan, S. (2023b). Zoochemical-mediated Nanoparticle Synthesis Using Marine Sponge Hyattella intestinalis (Lamarck, 1814) as a Reducing Agents. Uttar Pradesh Journal of Zoology, 44(18), 35– 41
- Kuruppuarachchi, S. U., & Gunathilake, V. K. (2022). Zoo-chemical profiling, in vivo toxicity and in vitro antiinflammatory properties of Luffariella herdmani marine sponge extract. J Adv Biotechnol Exp Ther, 5, 269-282.
- Muthiyan, R., Perumal, P., Muniswamy, K., Bhattacharya, D., Kundu, A., Sunder, J., & De, A. K. (2020). Zoo-chemical profile analysis and cytotoxicity screening of five marine sponge species collected from Andaman and Nicobar Islands.
- 7. Putra, M. Y., & Hadi, T. A. (2017). Chemical composition, antimicrobial, cytotoxic and antiplasmodial activities of three sponges from Buton Islands,

- Indonesia. *Indonesian Journal of Marine Sciences/Ilmu Kelautan*, 22(3).
- 8. Krishnan, G. S., Rajagopal, V., Joseph, S. R. A., Sebastian, D., Savarimuthu, I., Selvaraj, K. R. N., & Thobias, A. F. (2017). *In vitro, in silico* and *in vivo* antitumor activity of crude methanolic extract of *Tetilla dactyloidea* (Carter, 1869) on DEN Induced HCC in a Rat Model. *Biomedicine* & *Pharmacotherapy*, 95, 795-807.
- 9. Rivera, A. P., & Uy, M. M. (2012). In vitro antioxidant and cytotoxic activities of some marine sponges collected off misamis oriental coast, Philippines. *E-journal of Chemistry*, 9(1), 354-358.
- Tayone, J. C., Del Rosario, R. M., & Canencia, O. P. (2019). Extracts from egg strings of sea hare (*Dolabella auricularia*): yield, antioxidant activity, zoochemical profile, and toxicity. *International journal of advanced and applied sciences*, 6(1), 24-28.
- 11. Karnan R., Sukumaran M., Mariappan P., & Velavan S. (2023d). Zoo-mediated CuO Nanoparticles Synthesis Using Sea Urchin *Salmacis virgulata* (L. Agassiz & Desor, 1846) Test as a Novel Reducing Agent and Evaluated the In silico Antifungal Activity. *Uttar Pradesh journal of zoology*, 44(22), 154–167.
- Rompas, G., Lintang, R. A., Sumilat, D. A., Rumengan, I. F., Ginting, E. L., & Pangkey, H. D. (2022). Antibacterial Activity and Zoochemical Analysis of Sea Urchin *Diadema setosum* (Leske, 1778) Extract from Aertembaga Waters, Bitung City. *Jurnal Ilmiah PLATAX*, 10(2), 372-379.
- 13. Jayathilake, J. M. N., & Gunathilake, K. V. K. (2020). *In-vitro* anti-inflammatory activity, acute toxicity to zebrafish embryos and nutritional analysis of *Bohadschia vitiensis* water extract. *IJPSR*, Volume 11, Issue 9. 4501-4508.
- 14. Walag, A. M. P., Del Rosario, R. M., & Canencia, O. P. (2019). Zoochemical composition of selected sea stars collected from the Coastal Waters of Carmen, Agusan Del Norte, Philippines. Asian Journal of Biological and Life Sciences, 8(2), 53.
- Khattab, R. A., Elbandy, M., Lawrence, A., Paget, T., Rae-Rho, J., Binnaser, Y. S., & Ali, I. (2018). Extraction, identification and biological activities of saponins in sea cucumber Pearsonothuria graeffei. Combinatorial Chemistry &

- High Throughput Screening, 21(3), 222-231.
- Elbandy, M., Rho, J. R., & Afifi, R. (2014). Analysis of saponins as bioactive zoochemicals from the marine functional food sea cucumber *Bohadschia cousteaui*. European Food Research and Technology, 238, 937-955.
- Karnan, R., Sukumaran, M., Mariappan, P., & Velavan, S. (2023a). Insecticidal Effect of Zoochemicals Mediated Copper Oxide Nanoparticle Using Marine Sponge *Hyattella intestinalis* (Lamarck, 1814) and Molecular Docking. *Uttar Pradesh Journal of Zoology*, 44(15), 64–72.
- 18. Khalil, E. A., Swelim, H., El-Tantawi, H., Bakr, A. F., & Abdellatif, A. (2022). Characterization, cytotoxicity and antioxidant activity of sea urchins (*Diadema savignyi*) and jellyfish (Aurelia aurita) extracts. The Egyptian Journal of Aquatic Research, 48(4), 343-348.
- Nguyen, M. T., Tran, T. N., & Nguyen, H. V. (2022). Antibacterial and antioxidant activities of the extracts from marine snail *Hemifusus colosseus* (Lamarck, 1816). *Aquaculture, Aquarium, Conservation & Legislation*, 15(1), 251-260.
- Yahaya, N., Sakina, A. A., Haassan, A., & Muhammad, S. (2019). Zoochemical Screening and Antimicrobial Potential of Ground Beetle (Carabidae). Biochem Pharmacol (Los Angel), 8(265), 2167-0501
- 21. Laith, A. A., Ambak, M., Abol-Munafi, A. B., Nurhafizah, W. W. I., & Najiah, M. (2017). Metabolomic analysis of marine and mud crabs based on antibacterial activity. *Aquaculture Reports*, 7, 7-15.
- Umamageswari, P., R. Dineshkumar, P. Jayasingam, and P. Sampathkumar. "Antimicrobial activities of jellyfish, *Porpita porpita* from Southeast Coast of India." *Pharm. Biotechnol. Microbiol* 2016 (2016): 1-4.
- 23. Soundarapandian, P., Roy, S., & Varadharajan, D. (2014). Antioxidant activity in hard and soft shell crabs of *Charybdis lucifera* (Fabricius, 1798). *Journal of Aquaculture Research and Development*, 5.
- 24. George, I. E., Cherian, T., Ragavendran, C., Mohanraju, R., Dailah, H. G., Hassani, R., & Mohan, S. (2023). One-pot green synthesis of silver nanoparticles using brittle star *Ophiocoma scolopendrina*: Assessing biological potentialities of antibacterial, antioxidant,

- anti-diabetic and catalytic degradation of organic dyes. *Heliyon*, 9(3).
- 25. Hasaballah, A. I., El-Naggar, H. A., Abdelbary, S., Bashar, M. A., & Selim, T. A. (2022). Eco-friendly synthesis of zinc oxide nanoparticles by marine sponge, Spongia officinalis: antimicrobial and insecticidal activities against the mosquito vectors, Culex pipiens and Anopheles pharoensis. BioNanoScience, 1-16.
- 26. Sivanandham Velavan, R Shunmuga Vadivua, JJ. Vimala Sujia, B. Manimegalaia. (2020). Biosynthesis of silver nanoparticles using bivalvia shell extract and evaluation of its larvicidal activity. World Journal of Science and Research. 5(2): 12-16.
- Hamed, M. R., & Moradi, M. H. G. A. M. (2015). Biosynthesis of Silver Nanoparticles Using Marine Sponge. Oriental Journal of Chemistry, 31(4), 1961.
- Inbakandan, D., Sivaleela, G., Peter, D. M., Kiurbagaran, R., Venkatesan, R., & Khan, S. A. (2012). Marine sponge extract assisted biosynthesis of silver nanoparticles. *Materials Letters*, 87, 66-68.
- 29. Inbakandan, D., Venkatesan, R., & Khan, S. A. (2010). Biosynthesis of gold nanoparticles utilizing marine sponge *Acanthella elongata* (Dendy, 1905). *Colloids and Surfaces B: Biointerfaces*, 81(2), 634-639.