

Available online at http://www.harmanpublications.com

World Journal of Science and Research



ISSN: 2455 2208

Research Article Botany

PHYTOCHEMICAL SCREENING AND IN VITRO ANTI-OXIDANT ACTIVITY OF Justicia gendarussa LEAVES

M. Balakaviya and Dr. G. Santhi

PG and Research Department of Botany Kunthavai Naacchiyaar Govt. Arts College for Women (Autonomous) (Affiliated to Bharathidasan University), Thanjavur – 613 007, Tamil Nadu. India

ABSTRACT

The objective of the present study was evaluating the phytochemical constituents of *Justicia gendarussa* leaves extracts and their antioxidant activities. The different extracts were monitored for phytochemical screening. Total phenolic (279.00±13.85 mg/gm) and flavonoid (40.09±7.98 mg/gm) contents were measured in *Justicia gendarussa* leaves. The antioxidant potential of tested extracts was evaluated using total antioxidant capacity, ferrous ion and nitric oxide radical scavenging assays. This study demonstrated that, *Justicia gendarussa* leaves is a good source of natural antioxidants.

Keywords: *Hemidesmus indicus*, qualitative, quantitative and anti-stress.

INTRODUCTION

Antioxidants act as a defense mechanism that protects against oxidative damage, and include compounds to remove or repair damaged molecules. It can prevent/retard the oxidation caused by free radicals and sufficient intake of antioxidants is supposed to protect against diseases (Celiktar et al., 2007). Oxidative stress occurs when there is excessive free radical production and/or low antioxidant defense, which leads to chemical alterations of biomolecules causing structural and functional modification(Hoake and Pastorino, 2002).

Oxidative damage causing plays a significant pathological role in human diseases such as cancer, inflammation arthritis, diabetes and atherosclerosis (Halliwell, 1991). Recently there has been an upsurge of interest in the therapeutic potential of medicinal plants as antioxidants in reducing free radical induced tissue injury. Also many other plant species have been investigated in the search of novel antioxidants (Omar *et al.*, 2009),

Natural products contain different valuable chemical components such as phenolic compounds, phthalates, phenylpropanoids, terpenoids, essential oils, aromatic compounds, alkaloids, sterols, polysaccharides, fatty acids, anthocyanin, tannins, etc. (Oksman-Caldentey and Inze 2004; Picot et al. 2017; Mollica et al., 2015). They also have significant antioxidant activity (Embuscado 2015). Knowledge of these components in a plant not only helps for the quality control analysis of the plant but also manifests nature of the drug or formulation (Jain et al. 2011). To investigate the phytochemical screening and in vitro antioxidant activity of leaves Justicia gendarussa.

MATERIALS AND METHODS Collection of plant materials

The Justicia gendarussa were collected from Kdukaval, Thanjavur district, Tamil Nadu, India during March 2022. The collected leaves were washed several times with distilled water to remove the traces of impurities from the leaves. Then examined carefully, old infected and fungus damaged portion of the leaves were removed. Healthy

leaves were dried in room temperature and grind using grinder mixture. The powder was stored for further analysis

Preparation of plant extract

1 gram of dried powder of *Justicia* gendarussa leaves were transferred in to different conical flask (250ml). The conical flask containing 50ml of different solution such as ethanol and water. The conical flask containing *Justicia* gendarussa leaves were shaken well for 30 minutes by free hand. After 24 hrs, the extracts were filtered using Whatman filter paper No.1 and filtrate is used for further analysis.

Phytochemical screening

Chemical tests were carried out on the extract using standard procedures to identify the constituents as described by Sofowara (1993), Trease and Evans (1989) and Harborne (1973 and 1984). Total phenols estimated by the method of Edeoga *et al.*, (2005). Flavonoid determine by the method of Bohm and Kocipai-Abyazan (1994).

In vitro antioxidant activity

The antioxidant activity of extract was evaluated by the phosphomolybdenum method according to the procedure of Prieto *et al.* (1999). The chelating activity of the extracts for ferrous ions Fe²⁺ was measured according to the method of Dinis *et al.* (1994).

Nitric oxide radical scavenging activity was determined according to the method reported by Garrat (1964).

RESULTS AND DISCUSSION

Phytochemicals as well as medicinal plants, have remained the most abundant source of health care and life improvement since very long. Phytochemicals have great antioxidant potential and are of great interest due to their beneficial effects on health of human beings, and they give immense health benefits to the consumers. Epidemiological and animal trials suggest that the regular consumption of fruits and vegetables, and whole grains reduces the risk of various diseases linked with oxidative damage (Cieslik et al., 2006; Scalbert et al., 2005). In the present study was carried out on the Justicia gendarussa leaves extract revealed presence of medicinally the active constituents. Justicia gendarussa leaves extract showed that the presence of tannin, saponins, flavonoids, steroids, terpenoids, triterpenoids, polyphenol, glycoside, antroquinone and coumarins while alkaloids was absent in both aqueous and ethanolic extract. Significant amount of flavonoids (40.09±7.98 mg/gm) and total phenol (279.00±13.85 mg/gm) were observed in Justicia gendarussa leaves.

Table 1: Qualitative phytochemical analysis of Justicia gendarussa leaves extract

S. No	Phytochemicals	Ethanol extract	Aqueous extract
1	Tannin	++	+
2	Saponin	++	++
3	Flavonoids	++	+
4	Steroids	++	+
5	Terpenoids	++	+
6	Triterpeniods	++	+
7	Alkaloids	-	=
8	Antroquinone	+	+
9	Polyphenol	++	++
10	Glycosides	+	+
11	Coumarins	++	++

(+) Presence, (++) High concentrations and (-) Absences

Table 2: Quantitative estimation of Justicia gendarussa leaves powder

S. No	Phytochemicals	Results (mg/gm)	
1	Flavonoids	40.09±7.98	
2	Total phenol	279.00±13.85	

Values are expressed as mean \pm SD for triplicates

In vitro antioxidants activity

In the present study anti-oxidant activity of *Justicia gendarussa* leaves extract, *In vitro* antioxidant activity of the *Justicia gendarussa* extracts was observed by Total antioxidant capacity (TAC), Iron chelating antioxidant assay and Nitric oxide antioxidant activity methods. It was, however, observed that the 100 μg/ml extract possesses significant total antioxidant capacity

equivalent to ascorbic acid at higher concentration. The formation of the ferrozine– Fe^{2+} complex is interrupted in the presence of *Justicia gendarussa* leaves, indicating that have chelating activity with *Justicia gendarussa* leaves extract maximum observation at 100μ g/ml of inhibitions nearest to Std. (Ascorbic acid). The nitric oxide scavenging capacity was dose dependent with 100 to 500μ g/mL.

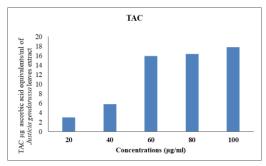
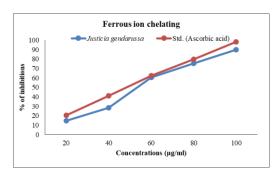


Figure 1: The total antioxidant capacity (TAC) of *Justicia gendarussa* leaves extract Figure 2: *In vitro* ant-oxidant ferrous ion



chelating of *Justicia gendarussa* leaves extract

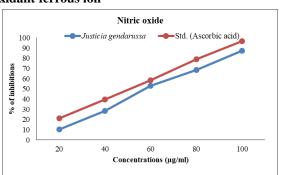


Figure 3: *In vitro* ant-oxidant Nitric oxide scavenging activity of *Justicia gendarussa* leaves

The total antioxidant capacity (TAC) was based on the reduction of Mo(VI) to Mo(V) by the extract and subsequent formation of green phosphate/Mo(V) complex at acid pH (Prieto *et al.* 1999). Ferrous iron can initiate lipid peroxidation by the Fenton reaction as well as accelerating peroxidation by decomposing lipid hydroperoxides into peroxyl and alkoxyl radicals (Halliwell, 1991; Fridovich, 1995).

REFERENCES

Bohm, B. A., & Kocipai-Abyazan, R. (1994). Flavonoids and condensed tannins from leaves of *Hawaiian vaccinium* and *V calycinium*. *Pacific Sci*, 48, 458-463.

Celiktas, O., Girgin, G. Ö. Z. D. E., Orhan, H. İ. L. M. İ., Wichers, H. J., Bedir, E. R. D. A. L., & Vardar-Sukan, F. (2007). Screening of free radical scavenging capacity and antioxidant activities of Rosmarinus officinalis extracts with focus on location and harvesting times. European Food Research and Technology, 224(4), 443-451.

Cieślik, E., Gręda, A., & Adamus, W. (2006). Contents of polyphenols in fruit and vegetables. *Food chemistry*, 94(1), 135-142.

CONCLUSION

This is a good study in which the authors have evaluated the *in-vitro* antioxidant activity of *Justicia gendarussa* leaves by TAC, ferrous ion and nitric oxide radical scavenging assay and the content of total phenolic and flavonoids. The antioxidant effect was plausibly due to the rich phytochemical contents.

Dinis, T. C., Cunha, A. P., & Almeida, L. (1994). Antioxidant activities of some extracts of Thymus zygis. *Free radical research*, 26(5), 469-478.

Edeoga, H. O., Okwu, D. E., & Mbaebie, B. O. (2005). Phytochemical constituents of some Nigerian medicinal plants. *African journal of biotechnology*, 4(7), 685-688.

Embuscado, M. E. (2015). Spices and herbs: Natural sources of antioxidants—a mini review. *Journal of functional foods*, 18, 811-819.

Fridovich, I. (1995). Superoxide radical and superoxide dismutases. *Annual review of biochemistry*, 64(1), 97-112.

Garrat, D. C. (1964). The Quantitative Analysis of Drugs. Quant. Anal. *Drugs*.

- Halliwell, B. (1991). Reactive oxygen species in living systems: source, biochemistry, and role in human disease. *The American journal of medicine*, 91(3), \$14-\$22.
- Harborne, J. B. (1973). *Phytochemical methods*, London. Chapman and Hall, Ltd. pp. 49-188.
- Harborne. J B. (1984). Phytochemical Methods.A Guide to Modern Technique of Plant analysis. London: *Chapman and Hall*, 78-210.
- Hoek, J. B., & Pastorino, J. G. (2002). Ethanol, oxidative stress, and cytokine-induced liver cell injury. *Alcohol*, 27(1), 63-68.
- Jain, V., Murugananthan, G., Deepak, M., Viswanatha, G. L., & Manohar, D. (2011). Isolation and standardization of various phytochemical constituents from methanolic extracts of fruit rinds of Punica granatum. Chinese Journal of Natural Medicines, 9(6), 414-420.
- Mollica, A., Costante, R., Akdemir, A., Carradori, S., Stefanucci, A., Macedonio, G., & Supuran, C. T. (2015). Exploring new Probenecid-based carbonic anhydrase inhibitors: Synthesis, biological evaluation and docking studies. *Bioorganic & Medicinal Chemistry*, 23(17), 5311-5318.
- Oksman-Caldentey, K. M., & Inzé, D. (2004).

 Plant cell factories in the postgenomic era: new ways to produce designer secondary metabolites. *Trends in plant science*, 9(9), 433-440.
- Omar, A. A., Hawazin, H. M., Asmita, V. P., & Gerald, B. (2009). Chemical composition and antioxidant activities of Jeddah corniche algae,

- Saudi Arabia, Saudi. J. Biol. Sci, 16, 23-29.
- Omar, A. H., Winker, D. M., Vaughan, M. A., Hu, Y., Trepte, C. R., Ferrare, R. A., & Liu, Z. (2009). The CALIPSO automated aerosol classification and lidar ratio selection algorithm. *Journal of Atmospheric and Oceanic Technology*, 26(10), 1994-2014.
- Picot, M. C., Zengin, G., Mollica, A., Stefanucci, A., Carradori, S., & Mahomoodally, M. (2017). In vitro and in silico studies of mangiferin from Aphloia theiformis on key enzymes linked to diabetes type 2 and associated complications. *Medicinal chemistry*, 13(7), 633-640.
- Prieto, P., Pineda, M., & Aguilar, M. (1999).

 Spectrophotometric quantitation of antioxidant capacity through the formation of a phosphomolybdenum complex: specific application to the determination of vitamin E. *Analytical biochemistry*, 269(2), 337-341.
- Scalbert, A., Johnson, I. T., & Saltmarsh, M. (2005). Polyphenols: antioxidants and beyond. *The American journal of clinical nutrition*, 81(1), 215S-217S.
- Sofowara, A. (1993). Medicinal plants and Traditional medicine in Africa. Spectrum Books Ltd, Ibadan, Nigeria, 191-289.
- Trease, G. E., & Evans, W. C. (1989).

 Pharmacognsy. 11th edn. Brailliar
 Tiridel can. Macmillian
 Publishers.U.S. (1984).

 Environmental protection Agency,
 Draft Criteria document for carbon
 tetrachloride, criteria and standards
 Division, office of Drinking,
 Washington, DC.